

Metadata of the article that will be visualized in OnlineFirst

ArticleTitle	Antidote activity of vitamin B ₁₂ derivative compared with its original and aqua forms; in vitro and in vivo study	
--------------	---	--

Article Sub-Title		
-------------------	--	--

Article CopyRight	Association of Food Scientists & Technologists (India) (This will be the copyright line in the final PDF)	
-------------------	--	--

Journal Name	Journal of Food Science and Technology	
--------------	--	--

Corresponding Author	FamilyName	Gromova
	Particle	
	Given Name	Olga A.
	Suffix	
	Division	
	Organization	Federal Research Center “Computer Science and Control” of Russian Academy of Sciences
	Address	Moscow, Russia
	Phone	
	Fax	
	Email	unesco.gromova@gmail.com
	URL	
	ORCID	http://orcid.org/0000-0002-7663-710X

Corresponding Author	FamilyName	Maiorova
	Particle	
	Given Name	Larissa A.
	Suffix	
	Division	
	Organization	Federal Research Center “Computer Science and Control” of Russian Academy of Sciences
	Address	Moscow, Russia
	Division	Institute of Macroheterocyclic Compounds
	Organization	Ivanovo State University of Chemistry and Technology
	Address	Ivanovo, Russia
	Phone	
	Fax	
	Email	maiorova.larissa@gmail.com
	URL	
	ORCID	http://orcid.org/0000-0003-3172-5621

Corresponding Author	FamilyName	Jafari
	Particle	
	Given Name	Seid Mahdi
	Suffix	
	Division	Department of Food Materials and Process Design Engineering
	Organization	Gorgan University of Agricultural Sciences and Natural Resources
	Address	Gorgan, Iran
	Division	Iran Food and Drug Administration
	Organization	Halal Research Center of IRI, Ministry of Health and Medical Education
	Address	Tehran, Iran
	Phone	
	Fax	
	Email	smjafari@gu.ac.ir
	URL	
	ORCID	http://orcid.org/0000-0001-6877-9549

Author	FamilyName	Salnikov
	Particle	
	Given Name	Denis S.
	Suffix	
	Division	Institute of Macroheterocyclic Compounds
	Organization	Ivanovo State University of Chemistry and Technology
	Address	Ivanovo, Russia
	Phone	
	Fax	
	Email	densal82@mail.ru
	URL	
	ORCID	http://orcid.org/0000-0002-1212-4428

Author	FamilyName	Torshin
	Particle	
	Given Name	Ivan Yu
	Suffix	
	Division	
	Organization	Federal Research Center "Computer Science and Control" of Russian Academy of Sciences
	Address	Moscow, Russia
	Phone	
	Fax	
	Email	tiy135@yahoo.com
	URL	
	ORCID	http://orcid.org/0000-0002-2659-7998

Author	FamilyName	Demidov
	Particle	
	Given Name	Vadim I.
	Suffix	
	Division	
	Organization	Ivanovo State Medical University, Ministry of Health of Russia
	Address	Ivanovo, Russia
	Phone	
	Fax	
	Email	13vid@mail.ru
	URL	
	ORCID	

Author	FamilyName	Tomilova
	Particle	
	Given Name	Irina K.
	Suffix	
	Division	
	Organization	Ivanovo State Medical University, Ministry of Health of Russia
	Address	Ivanovo, Russia
	Phone	
	Fax	
	Email	tomilovaivanovo@mail.ru
	URL	
	ORCID	

Author	FamilyName Particle Given Name Suffix Division Organization Address Phone Fax Email URL ORCID	Koifman O. I. Institute of Macrocyclic Compounds Ivanovo State University of Chemistry and Technology Ivanovo, Russia
Author	FamilyName Particle Given Name Suffix Division Organization Address Phone Fax Email URL ORCID	Kalacheva Alla G. Ivanovo State Medical University, Ministry of Health of Russia Ivanovo, Russia alla_kalacheva@mail.ru
Author	FamilyName Particle Given Name Suffix Division Organization Address Phone Fax Email URL ORCID	Bogacheva Tatiana E. Ivanovo State Medical University, Ministry of Health of Russia Ivanovo, Russia tatiana.boga4iova@yandex.ru
Author	FamilyName Particle Given Name Suffix Division Organization Address Phone Fax Email URL ORCID	Alexakhina Elena L. Ivanovo State Medical University, Ministry of Health of Russia Ivanovo, Russia alexakhina2013@yandex.ru
Author	FamilyName Particle Given Name Suffix Division Organization Address Phone Fax Email URL ORCID	Grishina Tatiana R. Ivanovo State Medical University, Ministry of Health of Russia Ivanovo, Russia farma37@bk.ru

Author	FamilyName Particle Given Name Suffix Division Organization Address Phone Fax Email URL ORCID	Gromov Andrei N. Federal Research Center “Computer Science and Control” of Russian Academy of Sciences Moscow, Russia gromlogin@gmail.com
Author	FamilyName Particle Given Name Suffix Division Organization Address Division Organization Address Phone Fax Email URL ORCID	Assadpour Elham Food Industry Research Co. Gorgan, Iran Food and Bio-Nanotech International Research Center (Fabiano) Gorgan University of Agricultural Sciences and Natural Resources Gorgan, Iran assadpour1170@gmail.com http://orcid.org/0000-0001-6667-0507
Author	FamilyName Particle Given Name Suffix Division Organization Address Division Organization Address Phone Fax Email URL ORCID	Ashaolu Tolulope J. Institute for Global Health Innovations Duy Tan University Da Nang, 55000, Viet Nam Faculty of Medicine Duy Tan University Da Nang, 55000, Viet Nam tolulopejoshuashaolu@duytan.edu.vn http://orcid.org/0000-0002-9397-6357
Schedule	Received Revised Accepted	 3 Feb 2025 13 Feb 2025
Abstract	<p>Vitamin B₁₂ (VB12) – essential nutrient, required for detoxication of homocysteine, support of the myelinization in neural tissue and of the hematopoiesis. Certain drugs (such as antibiotics or antituberculosis drugs) result in deep deficiency of VB12. Using VB12 and its derivatives as antidotes is a promising direction in pharmacology, that allows compensation of the toxic effects of the drugs by nutraceuticals. In the present work, interactions of isoniazid (IZ) (a toxic drug, used in the pharmacotherapy of tuberculosis) with various VB12 derivatives were studied. An in vitro study in aqueous solutions with different pH values showed that the hydrophobic derivative of VB12—heptamethyl ester of aquacyanocobyrinic acid (ACm) promoted oxidation of IZ and contributed to reducing its hepatotoxicity. The effects of ACm were compared with VB12 and aquacobalamin in a rat model of acute IZ-induced hepatitis. IZ intoxication resulted in higher levels of aspartate aminotransferase (AST). Administration of VB12 and ACm normalized AST levels; treatment with aquacobalamin or ACm normalized total protein levels in blood serum. ACm also attenuated bilirubin levels in the blood. All VB12 derivatives significantly reduced lipid peroxidation, which was increased after IZ model was reproduced. Histological analysis confirmed the protective effects of these compounds on the rats’ livers, kidneys, and brains: hepatocyte damage, inflammatory cell infiltration of liver tissues, acute ischemia of the renal cortex, and structural brain damage caused by IZ were all reduced. ACm had more positive effects on the liver than the other two compounds.</p>	
Keywords (separated by ' ')	Vitamin B ₁₂ - Hydrophobic derivatives - Antidotes - Antioxidant effect - Isoniazid - Neuroprotection - Hepatoprotection	
Footnote Information	O. I. Koifman: Deceased on 31/12/2023.	



Antidote activity of vitamin B₁₂ derivative compared with its original and aqua forms; in vitro and in vivo study

Olga A. Gromova¹ · Larissa A. Maiorova^{1,2} · Denis S. Salnikov² · Ivan Yu Torshin¹ · Vadim I. Demidov³ · Irina K. Tomilova³ · O. I. Koifman² · Alla G. Kalacheva³ · Tatiana E. Bogacheva³ · Elena L. Alexakhina³ · Tatiana R. Grishina³ · Andrei N. Gromov¹ · Elham Assadpour^{4,5} · Tolulope J. Ashaolu^{6,7} · Seid Mahdi Jafari^{8,9}

Revised: 3 February 2025 / Accepted: 13 February 2025
© Association of Food Scientists & Technologists (India) 2025

Abstract

Vitamin B₁₂ (VB12) – essential nutrient, required for detoxication of homocysteine, support of the myelinization in neural tissue and of the hematopoiesis. Certain drugs (such as antibiotics or antituberculosis drugs) result in deep deficiency of VB12. Using VB12 and its derivatives as antidotes is a promising direction in pharmacology, that allows compensation of the toxic effects of the drugs by nutraceuticals. In the present work, interactions of isoniazid (IZ) (a toxic drug, used in the pharmacotherapy of tuberculosis) with various VB12 derivatives were studied. An in vitro study in aqueous solutions with different pH values showed that the hydrophobic derivative of VB12—heptamethyl ester of aquacyanocobyrinic acid (ACm) promoted oxidation of IZ and contributed to reducing its hepatotoxicity. The effects of ACm were compared with VB12 and aquacobalamin in a rat model of acute IZ-induced hepatitis. IZ intoxication resulted in higher levels of aspartate aminotransferase (AST). Administration of VB12 and ACm normalized AST levels; treatment with aquacobalamin or ACm normalized total protein levels in blood serum. ACm also attenuated bilirubin levels in the blood. All VB12 derivatives significantly reduced lipid peroxidation, which was increased after IZ model was reproduced. Histological analysis confirmed the protective effects of these compounds on the rats' livers, kidneys, and brains: hepatocyte damage, inflammatory cell infiltration of liver tissues, acute ischemia of the renal cortex, and structural brain damage caused by IZ were all reduced. ACm had more positive effects on the liver than the other two compounds.

Keywords Vitamin B₁₂ · Hydrophobic derivatives · Antidotes · Antioxidant effect · Isoniazid · Neuroprotection · Hepatoprotection

Introduction

Insufficient supply of group B vitamins, especially vitamin B₁₂ (cyanocobalamin or VB12), is widespread in various populations. VB12 deficiency is especially common in people on a strict vegetarian diet (not taking special multivitamin complexes), in women of reproductive age (Oh et al. 2020), in patients receiving long-term antibacterial therapy (for example, in the treatment of tuberculosis), etc. toxic effects of the drugs can be counteracted by nutraceuticals such as VB12. A deficit vitamin B₁₂ in food or its low absorption in the stomach and intestines is associated with a severe form of hyperhomocysteinemia (Guéant et al.

2023), which leads to oxidative stress and lipid metabolism disorders, and provokes severe atherosclerosis (Haloul et al. 2020). Nutritional support with vitamin B₁₂ and folates through food supplements can significantly improve metabolism in patients with hyperhomocysteinemia. A clinical study showed that in severe hyperhomocysteinemia caused by nutritional or hereditary folate metabolism disorders cycle, deficit VB12 and folate increased cardiovascular risk and mortality (Levy et al. 2021). Supplementation with VB12 (1000 mcg/day) and folate (5000 mcg/day) for 12 weeks reduced level toxic metabolite—asymmetric dimethylarginine V plasma at patients With sharp ischemic stroke (Xia et al. 2014).

In tuberculosis, patients often have a combined deficiency of microelements, iron deficiency anemia, hypovitaminosis of group B vitamins, and a lack of antioxidants from vegetables. In a cohort study of various dietary patterns in 605

O. I. Koifman: Deceased on 31/12/2023.

Extended author information available on the last page of the article

55 patients with tuberculosis in China with a high risk of liver
56 damage caused by anti-tuberculosis drugs, a negative effect
57 of the dietary regimen "Offal meat, poultry, and vegetable oil
58 with insufficient vegetable consumption" on the indices of
59 cytolysis in the liver, the risk of damage, and liver dysfunction
60 was established. Therapy with antibiotics and specific
61 anti-tuberculosis drugs (isoniazid, metazid, larusan, strep-
62 tomycin, rifampicin) negatively interferes with the metabo-
63 lism of vitamins B₆ and B₁₂, and also leads to drug-induced
64 hepatitis (Gebremicael et al. 2019; Zhang et al. 2022). In the
65 treatment of hepatotoxic antibiotics, combined supplements
66 of spirulina enriched in particular with VB12 are tested
67 (Martin 2017). Correction of the diet with VB12, spirulina
68 and beta-carotene in rats with a tuberculosis model receiving
69 rifampicin and isoniazid led to a weakening of hepatotoxicity,
70 inflammatory blood markers and an improvement in the
71 immunohistochemistry of liver sections (Wang et al. 2024).
72 Replenishment of VB12 is very important for patients suffer-
73 ing from tuberculosis infection. They are recommended to
74 take additional VB12 in the form of food supplements, and if
75 necessary, in combination with other minor micronutrients
76 (vitamin B₆, vitamin A, beta-carotene) while maintaining
77 sufficient consumption of meat by-products, poultry, fruits
78 and vegetables.

79 Antidotes are chemicals that alter the action of a poison
80 in the body to prevent, reverse, or mitigate the toxic effects.
81 Examples of mechanisms by which antidotes work include
82 competition at a receptor site, alteration of a metabolic pro-
83 cess, engaging a counter-regulatory physiologic process, or
84 hastening the excretion or detoxification of a toxin. The stud-
85 ies dealing with vitamins and their derivatives as possible
86 antidotes are quite few. An important area of research in
87 pharmacology and physico-chemical medicine of vitamin
88 derivatives is the search for antidotes against targeted phar-
89 maceuticals. Aquacobalamin (aqua form of vitamin B₁₂) is
90 recommended as an antidote at cyanide poisoning. The stud-
91 ies of its derivative, diaquacobinamide, has showed that it
92 is notably more effective than aquacobalamin (AQ) at the
93 poisoning (Lee et al. 2016). Moreover, it could potentially
94 serve as a methyl mercaptan (Philipopoulos et al. 2022) and
95 hydrogen sulfide (Jiang et al. 2016) antidote.

96 In general, the establishment of the mechanisms for anti-
97 dote action of vitamin B₁₂ (VB12) and its derivatives is an
98 interesting and important direction in the molecular pharma-
99 cology. The known biological roles of cobalamins (includ-
100 ing cyanocobalamin, known as VB12) and the possibility
101 of their chemical modifications make it possible to develop
102 new drugs. In particular, a promising area of research is the
103 search for cobalamin/corrin antidotes (Hendry-Hofer et al.
104 2021), including those against intoxication with pharmaco-
105 logical drugs. Micro- and nanoencapsulation technologies
106 are important for targeted delivery and modulation of the
107 biological properties of such molecules (Gromova et al.

2021). Also, there are compounds with hydrophobic modi- 108
fications of the corrin ring of VB12 – for example, aquacya- 109
nocobyric acid heptamethyl ester (ACm). 110

111 Earlier we have shown that ACm, VB12, and AQ can
112 be recommended for further study as analgesics and anti-
113 inflammatory agents (Gromova et al. 2021). The ACm, in
114 particular, is soluble in water and a wide range of organic
115 solvents and exhibits a biological activity (protection of the
116 myelin sheath of neurons) under conditions of toxic stress
117 (thiosemicarbazide) (Gromova et al. 2022). The introduc-
118 tion of currently registered forms of VB12 and AQ saved
119 50% of the lives, and the introduction of unregistered ACm
120 saved 33% of the animals. In vitro studies were carried out
121 to explain these important and unexpected results for ACm.
122 Analysis of the electronic absorption spectra indicated the
123 possibility of direct interaction of the toxic thiosemicar-
124 bazide with AQ. However, the binding constant of the sub-
125 stance was quite small, which did not allow for explaining
126 the observed positive effect of AQ on reducing the toxic-
127 ity of thiosemicarbazide due to this interaction. The bind-
128 ing constant of this toxicant to the ACm was close to zero.
129 Therefore, the establishment of the mechanisms of the anti-
130 dote action of these compounds, especially ACm, requires
131 the involvement of additional fundamental research in vitro.

132 The interaction mechanisms between the studied VB12
133 derivatives in aqueous media and the toxicant molecules
134 might be very complex. For example, substances with sulfur-
135 containing functional groups (thiols, sulfites, as in thiosemi-
136 carbazide) and/or nitrogen-containing groups (amino acids,
137 pyridine, as in thiosemicarbazide or isoniazid (IZ)) can
138 directly interact with the cobalt nucleus of VB12 hydropho-
139 bic derivatives containing water in axial position, as axial
140 ligands. Then, such substances will replace the water mol-
141 ecule and form complexes with derivatives. Such complexes
142 can undergo further chemical transformations stimulated by
143 the cobalt ion as a kind of catalytic center. VB12 and its
144 derivatives, together with many other important biomol-
145 ecules in supramolecular assemblies, possess an impressive
146 variety of functional properties which are used in natural
147 systems performing their vital functions in living organisms.
148 Previously, we successfully encapsulated VB12 in nanoen-
149 gineered polymer capsules. The formation of molecular
150 assemblies at the interfaces is a specific feature of this class
151 of compounds (Valkova et al. 2002, 1999, 1996a; Vu et al.
152 2016; Maiorova et al. 2018). Self-assembly is a key player
153 in materials nano-architectonics (Ariga et al. 2019, 2017,
154 2021; Webre et al. 2018; Petrova et al. 2014; Oldacre et al.
155 2017; Brenner et al. 2017). Supramolecular polymers have
156 been created using diverse self-assembly strategies wherein
157 biomolecules are employed (Shee et al. 2020; Stulz 2017).
158 The possibility of the self-assembly of compounds into 2D
159 and 3D nanostructures possessing controlled properties
160 was demonstrated (Valkova et al. 1997, 1996b; Maiorova

et al. 2015; Kharitonova et al. 2018). Similar mono- and heteromolecular nanostructures containing drugs can be prepared in vitro (Soares et al. 2018; Nowak et al. 2020; Zeytunluoglu and Arslan 2022; Ouyang et al. 2019; Moraes Profirio et al. 2018) with a subsequent introduction into the body for a therapeutic effect or form spontaneously when molecular solutions were introduced. Recently, the formation of supermolecular nanoentities (SMEs) of VB12 derivative (viz. heptabutyl ester of aquacyanocobyrinic acid, ACBuCby) have been reported, i.e., unique nanoparticles exhibiting strong non-covalent intermolecular interactions and possessing intriguing properties (Maiorova et al. 2023). Besides reproducing the functional properties of VB12 complexes with proteins in living organisms, the nanoparticles demonstrate important advantages over VB12. They are more effective in oxygen reduction/evolution reactions and in transformations into other forms (Maiorova et al. 2023). Such nanoparticles can become an alternative form of VB12. Also, the first example of the formation of nanoparticles of ACm in protein nanocarriers and neuroprotective activity in vivo of the own nanoform of the drug has been revealed (Maiorova et al. 2024).

IZ is one of the first synthetic drugs against *Mycobacterium tuberculosis* and the first line of pharmacotherapy for tuberculosis (Vilchèze and Jacobs 2019). A significant disadvantage of IZ is its high hepatotoxicity (Erwin et al. 2019). Among animals, IZ intoxication in dogs is most severe (ASPCA Animal Poison Control Center Phone 2022). In case of IZ poisoning, pyridoxine hydrochloride (vitamin B₆) is used in doses equivalent to the dose of IZ taken (~ 50 mg/kg pyridoxine intravenously) (Villar et al. 1995). The high toxicity of IZ limits its use in patients with (i) liver disease, (ii) a tendency to convulsions, and (iii) low levels of pyridoxine in the blood (because of the formation of the IZ-pyridoxine complex) (O'Connor and Isoniazid 2022). Acute IZ poisoning leads to recurrent generalized tonic-clonic seizures, severe metabolic acidosis, liver and kidney damage, hematological disorders (Sridhar et al. 2012), mitochondrial insufficiency (Zhang et al. 2021), anorexia, limb tremor, and coagulopathy (Edgar et al. 2022). By long-term treatment of pulmonary forms of tuberculosis with IZ, toxic optic neuropathy of the optic nerve (Orssaud et al. 2022), psychoses and other neuropsychiatric disorders can develop (Gomes et al. 2019).

Previously, we have shown that certain hydrophilic derivatives of VB12 can be effective oxidizers of IZ and its toxic metabolites (Tumakov et al. 2019). Therefore, it is of interest to study the interactions of the toxicant IZ with ACm. Here the influence of VB12 semi-synthetic derivative on the effects associated with taking isoniazid, an anti-tuberculosis drug, was studied. We present: (1) molecular mechanisms of interaction between ACm and IZ, taking into account the pH of the solution; (2) the results of the hepato-, neuro- and

nephroprotective effects of ACm in comparison with VB12 and AQ in a model of acute hepatitis caused by IZ in rats. The effects of IZ when interacting with various VB12 derivatives were studied in vitro (in aqueous solutions) and in vivo (in a rat model of acute IZ-induced hepatitis with biochemical and histological tests).

Materials and methods

IZ (98%, from Alfa Aesar) was used without further purification. Organic solvents were purchased from Sigma-Aldrich. Acetate (0.01 M, pH = 4.7), phosphate (0.01 M, pH = 7.2), borate (0.01 M, pH = 9.2), and carbonate (0.01 M, pH = 10.7) buffers were used to control the pH as required.

Synthesis of ACm

Dicyanocobyrinic acid heptamethyl ester was produced and purified following the known procedure (Gromova et al. 2022). The structure (Fig. 1b) of this chemical was confirmed by MALDI-TOF mass spectroscopy. ACm was prepared by adding CH₃COOH to dicyanocobyrinic acid heptamethyl ester in ethanol/water (70:30) solution to pH = 4 and then passing a stream of nitrogen through the solution for ca. 24 h, as previously described (Gromova et al. 2022).

Photophysical characterization

VB12 was characterized by electron absorption spectra recorded with Shimadzu-UV-1800 and UV – Vis Cary 60 spectrophotometers.

Animal research

The study was carried out on 50 white male rats weighing 200–300 g in accordance with “Rules of good laboratory practice” (Appendix to the order of the Ministry of Health of the Russian Federation No. 199n dated 04/01/2016) (Dale et al. 2018; Brown et al. 2017) and allowed by the local ethical committee of IvGMA. During the studies, animals were kept under standard conditions in accordance with Directive 2010/63/EU of the European Parliament and of the Council of the European Union of 22 September 2010 concerning the protection of animals used in scientific studies. Indoor air control was in compliance with environmental parameters (temperature 18–26 °C, humidity 46–65%). The rats were kept in standard plastic cages with bedding; the cages were covered with steel lattice covers with a stern recess. The floor area per animal met regulatory standards. The animals were fed in accordance with Directive 2010/63/EU. The animals were given water ad libitum. The water was purified and normalized

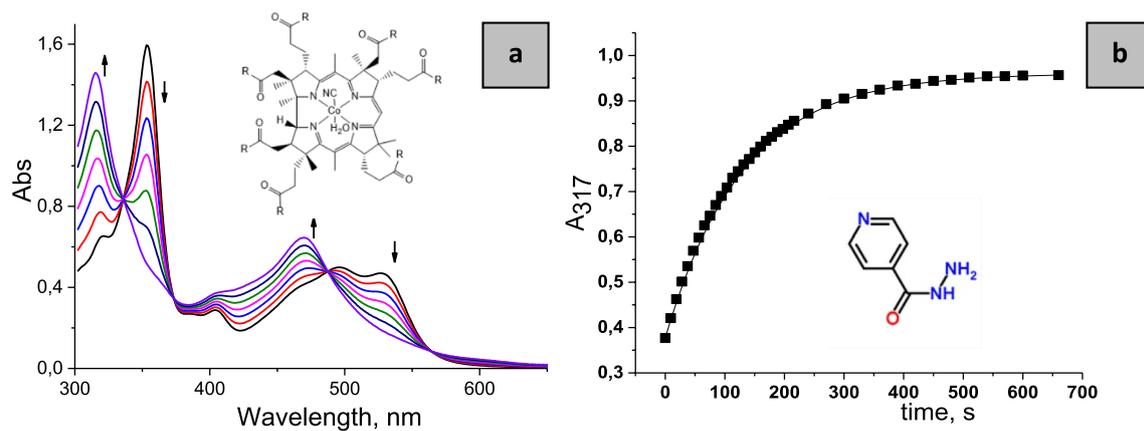


Fig. 1 **a** Spectral changes for the interaction of IZ with ACm in water under anaerobic conditions. **b** Kinetic curve for the reduction of Co^{3+} to Co^{2+} in ACm with IZ within aqueous solution. $[\text{ACm}]_0 = 5 \times$

10^{-5} M; $[\text{IZ}] = 1 \times 10^{-2}$ M; $\text{pH} = 7.4$; 25°C . The inserts show aquacyanocobyrinic acid heptamethyl ester (ACm), $\text{R} = \text{OCH}_3$ (**a**) and IZ (**b**)

258 for organoleptic properties in terms of pH, dry residue,
259 reducing substances, carbon dioxide, nitrates and nitrites,
260 ammonia, chlorides, sulfates, calcium and heavy metals in
261 standard drinkers with steel spout lids.

262 Animals were divided into 5 groups: the 1st group ($n = 10$)—intact control; in the 2nd, 3rd, 4th and 5th groups
263 of animals, the model of acute hepatitis was reproduced
264 by intragastric administration of IZ hepatotoxin at a dose
265 of 540 mg/kg body weight per day for 6 days (Couto and
266 Cates 2019); the 3rd group of animals ($n = 10$) was intra-
267 muscularly injected with a solution of 0.5 mg VB12/ml at a
268 dose of 60 $\mu\text{g}/\text{kg}$ of animal weight per day simultaneously
269 with IZ for 6 days and then another 10 days (registration
270 number P No. 015993/01, OJSC "Borisovskiy Factory of
271 Medical Preparations", Republic of Belarus); animals of
272 the 4th group were injected with AQ at a dose of 60 $\mu\text{g}/$
273 kg body weight per day intramuscularly (according to the
274 same scheme as in group 3); in the 5th group of rats ($n = 10$), ACm in water was administered intramuscularly
275 (according to the same scheme as in group 3) at a dose of
276 60 $\mu\text{g}/\text{kg}$ animal weight per day.

277 On the 17th day of the study, blood was taken for bio-
278 chemical studies and sectional material of the liver, kidneys,
279 and brain was taken for histopathological examination.
280 Aspartate aminotransferase (AST) and alanine aminotrans-
281 ferase (ALT) activity, levels of total protein, direct and total
282 bilirubin (using Olvex standard kits), and malondialdehyde
283 (MDA) were determined in the blood by the Jagi method
284 (Morris et al. 2022). In the comparison groups, the process
285 intensity of lipid peroxidation (LPO) in blood serum was
286 determined by the method of induced chemiluminescence.
287 Statistical data processing was carried out using the Statistica-10 program and Excel spreadsheet packages; the differences were evaluated using Mann–Whitney U-test at the upper level of significance $P < 0.05$.

Histological analysis

293 On the 17th day, the tissue sections of brain were prepared and
294 fixed in 10% neutral formalin solution; one day later, the area
295 of the precentral gyrus of the forebrain, cerebellum, and brain
296 stem were isolated using frontal incisions. After evisceration,
297 the liver and kidneys were fixed in a 10% neutral formalin
298 solution; after 1 day, the organs were dissected, fragments of
299 the right and left lobes of the liver, cortical sections of the
300 right and left kidneys were isolated and re-fixed. After the
301 secondary fixation and washing of the material, the dehydration
302 of the tissues of the brain, liver and kidneys was carried
303 out using 99% isopropyl alcohol. Subsequently, tissue sam-
304 ples were embedded in paraffin. Histological Sections 5–6
305 μm thick were made on a sledge microtome "Microm" and then
306 stained with hematoxylin and eosin. Duplicate sections of the
307 liver and kidneys were stained with Schiff's reagent, the brain
308 with toluidine blue according to the Nissl method. The assess-
309 ment of pathological changes in the organs of rats when mod-
310 eling toxic damage took into account the degree of circulatory
311 disorders, the characteristics of the inflammatory response,
312 and structural changes in parenchymal elements. Micropho-
313 tographs were obtained using a microscope "Micros" MS-200
314 and a digital eyepiece camera DCM 900. The degree of patho-
315 logical changes in each photo was estimated by experts in a
316 1–5 points score (5 – most pronounced histological damage,
317 1 – no histological damage). The statistical difference in scores
318 between the groups was estimated by Mann–Whitney U test as
319 significance level of $P < 0.05$.
320

321 **Results and discussion**322 **Interaction of ACm with isoniazid in aqueous media**
323 **(in vitro results)**

324 Compared to VB12, in the structure of ACm, there is no
325 nucleotide base, and seven amide side chains are replaced
326 by the ester groups (Fig. 1a). It was found that ACm can act
327 as an effective oxidizer of IZ. During the reaction, Co^{3+} ion
328 of ACm is reduced to Co^{2+} . Figure 1a shows the spectrum
329 changes corresponding to this process. A typical kinetic
330 curve for this reaction is shown in Fig. 1b.

331 It has been established that the kinetic curve can be well
332 linearized in the coordinates " $\ln(A_\infty - A)$ —time", which indi-
333 cates the first order of the reaction (estimated by ACm). The
334 observed ACm recovery rate constant *versus* IZ concentra-
335 tion (Fig. 2a) shows that the value of reduction rate constant
336 for ACm increases linearly with higher levels of IZ. There-
337 fore, the order for IZ is equal to one, *i.e.* the reaction rate
338 is related to the reactant concentrations as follows (Eq. 1):

$$339 \quad r = k \times [\text{ACm}]_0 \times [\text{IZ}]_0 \quad (1)$$

340 where $[\text{ACm}]_0$ is the total concentration of ACm and $[\text{IZ}]_0$ is
341 the total concentration of isoniazid (in an aqueous solution).

342 It was found that the spectra of the initial and IZ-reduced
343 ACm reactions with IZ do not change in the pH range from
344 3.2 to 8.5. But at higher pH values, the rate of reduction of
345 ACm with IZ increases (Fig. 2b). IZ has acid–base proper-
346 ties and exists in solution in four forms with $\text{pK}_{a1} = 1.99$,
347 $\text{pK}_{a2} = 3.67$, $\text{pK}_{a3} = 10.89$ (Fig. 3). In an acidic medium,
348 protonation of the nitrogen atoms of the pyridine (pK_{a1})
349 and hydrazide (pK_{a2}) fragments is possible; in an alkali-
350 ne medium, deprotonation of one nitrogen atom in the
351 hydrazide fragment (pK_{a3}) can occur.

352 ACm transforms into its hydroxo form only in an alkaline
353 medium (at $\text{pH} > 10$). Since an increase in the reduction
354 rate of ACm observed at a significantly lower pH, the data
355 obtained can be explained by the participation of both pro-
356 tonated and deprotonated forms of IZ in the process under
357 study. The rate increase also shows that the deprotonated
358

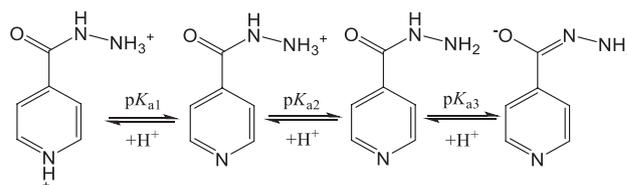


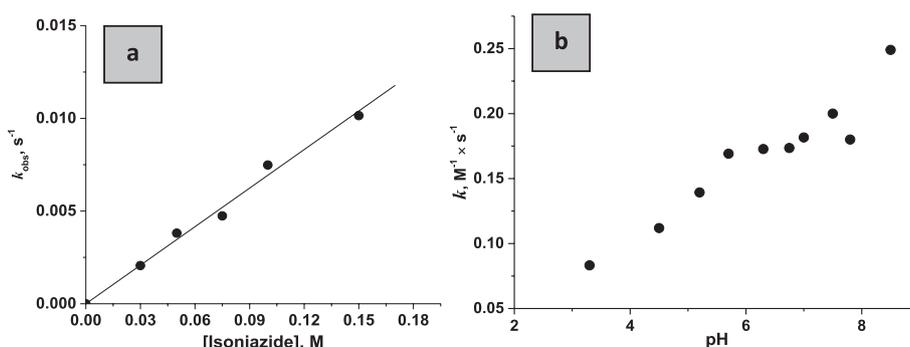
Fig. 3 Acid–base properties of isoniazid

359 forms of IZ are stronger reducing agents. According to
360 the pK_a values of IZ, at physiological pH, it should be in
361 a neutral form. Our data show at that pH value, the oxida-
362 tion of IZ by ACm does proceed. Thus, at a pH close to
363 physiological, the ACm oxidizes IZ. The reaction products,
364 similarly to what was shown in our previous work for cobi-
365 namide (Tumakov et al. 2019), are the reduced derivative of
366 ACm containing Co^{2+} and the oxidation products of IZ: *i.e.*,
367 isonicotinic acid, isonicotinamide, and pyridine-4-carbox-
368 aldehyde, which do not exhibit hepatotoxic properties. The
369 presence of isonicotinic acid in the reaction products sug-
370 gests that ACm can oxidize the radical forms of IZ, which
371 are responsible for its hepatotoxicity.

372 The results of acute isoniazid poisoning in rats

373 Preliminary experiments with IZ showed that when it was
374 administered at a dose of LD_{50} , intoxication occurred no
375 later than the first day from the moment of administration;
376 then the animals gradually recovered from the state of visu-
377 ally determined intoxication, and there was no lethality at
378 later periods of observation. Repeated administration of IZ
379 in LD_{50} dose after 24 h to rats (acute poisoning) resulted in
380 100% lethality. Following the literature, daily administra-
381 tion of IZ at a dose of 600 mg/kg (1/2 of LD_{50}) leads to
382 a 20% lethality of rats only by the end of the first week of
383 administration (on days 6–7 of administration); and only
384 by the end of 3 weeks, the mortality can reach 100%. This
385 phenomenon can be regarded as a true sign of the develop-
386 ment of tolerance in rats to IZ, since over the entire period of
387 the experiment, animals can withstand exposure to 10 LD_{50}
388 doses of IZ (Badrinath et al. 2022).

Fig. 2 **a** Rate constant of ACm reduction by IZ *versus* its concentration (conditions identical to those presented in Fig. 1). **b** Rate constant of ACm reduction by IZ *versus* pH. $[\text{ACm}]_0 = 5 \times 10^{-5} \text{ M}$, 25°C



389 The introduction of a high dose of IZ for 6 days led to
390 significant liver damage: when compared with intact control
391 (group 1), in animals of the 2nd group (which was sub-
392 sequently used as the main comparison group), a signifi-
393 cant increase in the level of blood AST (from 0.36 to 0.73
394 mM/h/l) and direct bilirubin (from 8.5 to 18.4 μ M/l) was
395 found. In addition, there was a decrease in the levels of total
396 serum protein (from 56.2 to 45.6 g/l), which corresponds
397 to a decrease in the protein-synthetic function of the liver.
398 A well-known feature of the IZ model of hepatotoxicity
399 is that there is no significant increase in malondialdehyde
400 (MDA) levels in liver injury. Indeed, there was no significant
401 changes in MDA levels when the model was reproduced
402 (Table 1).

403 These biochemical parameters (Table 1) reflect the func-
404 tional state of the liver. The levels of AST and ALT in the
405 blood are enzymes-indicators of cytolysis (primarily, of
406 hepatocytes). Enzymatic AST and ALT are present in sig-
407 nificant amounts in the liver and kidneys and, therefore, their
408 concentrations in the blood are normally low. The marker
409 of excretory and antitoxic function of the liver is bilirubin,
410 one of the intermediate products of hemoglobin breakdown
411 occurring in hepatocytes (Fig. 4). MDA is formed during
412 the degradation of fats and is a marker of oxidative stress.
413 During the cytolysis of hepatocytes, losses of MDA, bilirubin,
414 AST, and ALT enzymes occur, and the levels of these
415 biochemical markers in the blood increase. Changes in the
416 level of total protein are a sign of a gross pathology of the
417 liver and a violation of its synthetic function. According to
418 Mann–Whitney U-test, the reproduction of the IZ model led
419 to a significant increase of AST levels in blood. Cyancobala-
420 min application resulted in a significant decrease of AST (P
421 < 0.05). Reproduction of the IZ model led to a significant
422 decrease of total protein and application of ACm resulted
423 in an increase of total protein towards the original levels (P
424 < 0.05).

425 Changes in markers of liver and kidney function were
426 confirmed by the results of histological analysis (Fig. 5). In

intact animals (group 1), the microscopic image of the liver
tissue corresponded to the norm in all tissue samples. Within
a single hepatic lobule, while maintaining histoarchitecton-
ics, uniform perfusion of sinusoids was observed both in the
central and periportal zones of the lobule. Hepatocytes had
a normal configuration and uniform coloring with a correct
distribution of ultrastructures in the cytoplasm (Fig. 5A).
There were single lymphocytes in the stroma of the portal
tracts (Fig. 5B). The study of the cortical zone of the kid-
neys showed the normal structure of the glomeruli with a
physiological level of perfusion and free mesangial space.
Nephrocytes within the convoluted proximal and distal
tubules were also undamaged (Fig. 5C). The brain of rats in
the control group had a normal level of perfusion without
signs of aggregation of erythrocytes in the capillaries and
swelling of the nervous tissue; the neurons were character-
ized by a normal shape and size with clear contours of the
nuclei. Nissl lumps were evenly distributed in the cytoplasm
of pyramidal neurons in the forebrain cortex (Fig. 5D).

When reproducing the IZ model (group 2), negative
changes in the liver tissue were revealed in all of the rel-
evant samples. Against the background of anemia in the
central veins and sinusoids, widespread vacuolar dystro-
phy of hepatocytes, intracellular cholestasis, and discom-
plexation of the hepatic beams in the central zone of the
hepatic lobules were noted (Fig. 5F). In the 1st, 2nd and 4th
samples, there was focal necrosis of hepatocytes (Fig. 5E).
Pronounced lymphohistiocytic infiltration of the stroma
of the portal tracts with spread to the stroma of the sinu-
soids; eosinophils were present in the inflammatory infil-
trate (Fig. 5I). When examining the kidneys, anemia of the
glomeruli was noted against the background of spasm in
the interlobular arteries (Fig. 5J), an accumulation of a pro-
tein substance in the mesangial space. Nephrocytes of the
proximal convoluted tubules underwent vacuolar degenera-
tion, and PAS-negative masses were noted in the lumens of
the tubules (Fig. 5G). In the brain, against the background
of spasm in small-caliber arteries, there was a pronounced

Table 1 Biochemical parameters of blood serum in the comparison groups

Group No	Observation groups	Biochemical indicators				
		AST (mM/h/l)	ALT (mM/h/l)	Total serum protein (g/l)	MDA (nM/ml)	Bilirubin direct (μ M/l)
1	Control intact	0.36 \pm 0.17	0.45 \pm 0.10	56.2 \pm 3.1	3.91 \pm 0.43	8.5 \pm 4.5
2	IZ, control	0.73 \pm 0.13 ^a	0.58 \pm 0.33	45.6 \pm 3.4 ^a	4.72 \pm 1.34	18.4 \pm 6.3 ^a
3	IZ + VB12	0.58 \pm 0.05 ^b	0.37 \pm 0.12	47.6 \pm 2.8	4.22 \pm 0.93	16.0 \pm 7.8
4	IZ + AQ	0.64 \pm 0.13	0.39 \pm 0.13	49.0 \pm 1.2 ^b	4.13 \pm 0.86	15.0 \pm 7.6
5	IZ + ACm	0.61 \pm 0.10 ^b	0.42 \pm 0.11	53.8 \pm 1.5 ^{b,c}	4.13 \pm 0.54	9.7 \pm 5.1 ^{b,c}

Note: Data are in the format of mean \pm SD (standard deviation). Significant differences were noted: ^abetween intact (group 1) and model (group 2); ^bbetween the control group (group 2, IZ) and comparison groups (3, 4, 5); ^cbetween VB12 (group 3) and groups 4, 5 (Mann–Whitney test): $P < 0.05$

ACm aquacyanocobyrinic acid heptamethyl ester

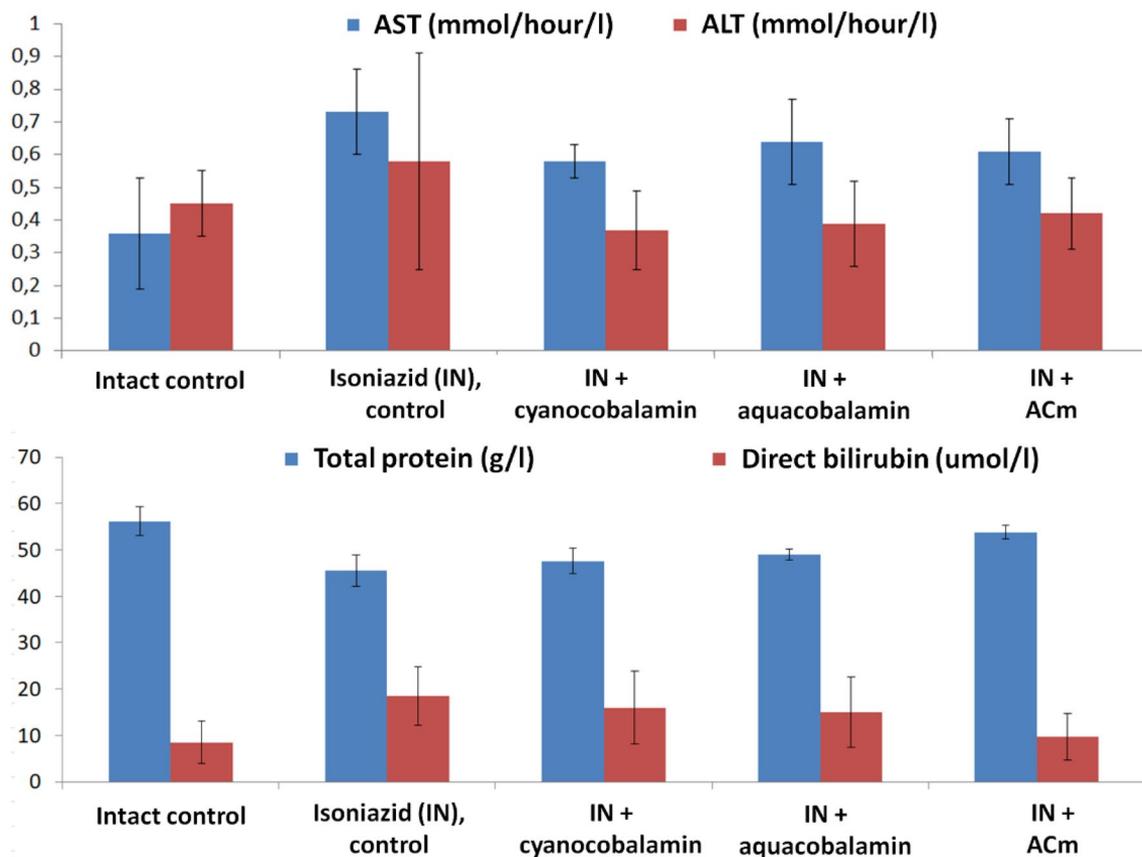


Fig. 4 Changes in biochemical markers of liver and kidney function with the use of VB12 derivatives

465 plethora of pial and intracerebral veins, stasis of erythrocytes
 466 in capillaries, perivascular and pericellular edema of the
 467 nervous tissue (Fig. 5K). There were small focal diapedetic
 468 hemorrhages in the cortex of the forebrain hemispheres.
 469 Changes in the pyramidal neurons of the forebrain and piri-
 470 form neurons of the cerebellum were focal in nature with an
 471 ischemic type of damage. Pycnosis was observed along with
 472 hyperchromia of the cytoplasm in cortical neurons with axon
 473 swelling (Fig. 5H); similar changes were found in the cere-
 474 bellum in single pear-shaped neurons. Irreversible changes
 475 in neurons, accompanied by plasmorhexis and activation of
 476 the neurophagic reaction of glial cells, were found in the
 477 1st and 5th samples (Fig. 5L). The integral expert score
 478 of histological damage was 3.2 ± 1.5 for IZ model and 1.3
 479 ± 0.7 for intact group which was significant according to the
 480 Mann–Whitney U-test ($P < 0.05$).

481 In general, the toxic effects of IZ had morphological
 482 confirmation. In the liver, there was ischemia of the centers
 483 of the lobules, subtotal protein (vacuole) dystrophy of
 484 hepatocytes (which corresponds to a decrease in the levels of
 485 total protein in a biochemical blood test, Table 1), impaired
 486 conjugation of bilirubin (with an increase in the level of
 487 direct bilirubin in the blood, Table 1), and inflammatory

488 cell infiltration stroma of the liver with the presence of
 489 eosinophils in the infiltrate (a sign of drug-induced hepatis-
 490 tis). In the kidneys, intoxication with IZ was accompanied
 491 by severe anemia of the cortex, development of widespread
 492 protein dystrophy of nephrocytes in the proximal convo-
 493 luted tubules, damage to the glomerular filter with exces-
 494 sive filtration of protein compounds (also contributing to the
 495 development of hypoproteinemia, Table 1). In the brain, IZ
 496 stimulated circulatory disorders (mainly in the microcircula-
 497 tory bed) with the development of moderately pronounced
 498 edema of the nervous tissue and with focal (mostly revers-
 499 ible) damage to the neurons of the cortex in the forebrain
 500 and cerebellum.

Biochemical effects of the studied compounds

501
 502 The three VB12 derivatives showed significant differences
 503 in the profile of action on the studied biochemical mark-
 504 ers. In the 3rd (VB12) and 5th (ACm) groups, a significant
 505 decrease in AST levels was noted compared with the IZ-only
 506 control (from 0.73 to 0.58...0.61 mM/h/l, Table 1). In the
 507 4th (AQ) and 5th (ACm) groups, the content of total protein
 508 in the blood serum increased significantly compared with the

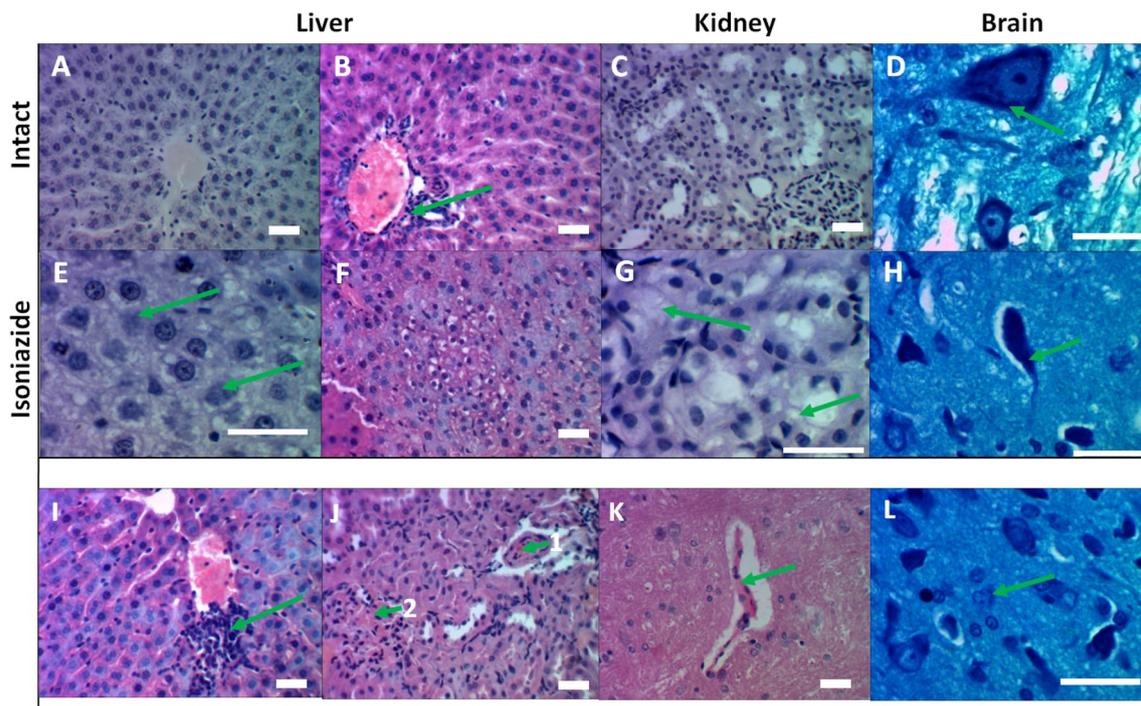


Fig. 5 Histological images of the liver, kidneys and brain in the reproduction of the model of IZ hepatotoxicity. The intact control (A–D) and the IZ model (E–L). Staining with PAS reaction (A, C, E, G), hematoxylin and eosin (B, F, I, J, K), toluidine blue according to Nissl (D, H, L). Magnification $\times 480$ (A, B, C, F, I, J, K) and $\times 1200$ (D, E, G, H, L), scale bar corresponds to 100 μm . A The structure of an unchanged hepatic lobule with a trabecular arrangement of hepatocytes. B Single lymphocytes in the stroma of the portal tract against the background of plethora of the portal vein. C The renal glomerulus has a capsule lumen and a free mesangium, tubular epithelial cells with a homogeneous color of the cytoplasm. D A pyrami-

dal neuron of a habitual configuration with a uniform distribution of Nissl clumps in the cytoplasm. E Vacuolar dystrophy, focal necrosis of hepatocytes in the stage of cytolysis. F Vacuolar degeneration of hepatocytes with impaired histoarchitectonics of the hepatic lobule. G Vacuolar degeneration of nephrocytes of the proximal convoluted tubules. H Pycnomorphic pyramidal neuron with swollen axon. I Lymphohistiocytic infiltrate with single eosinophils of the periportal zone. J Spasm of the interlobular artery, stromal edema (1), eosinophilic masses in the mesangium of the glomerulus (2). K Hemostasis in the capillary, perivascular and pericellular edema of the nervous tissue. L The reaction of microglia in the area of the dead neuron

509 IZ control, which indicates a possible improvement in the
510 protein-synthetic function of the liver. The introduction of
511 ACm led to the normalization of the AST and protein levels
512 in the blood, as well as to the normalization of the direct
513 bilirubin level in the blood (decrease from 18.4 to 9.7 $\mu\text{M/l}$).

514 The intensity results of the process of LPO by the chemi-
515 luminescence method confirmed the conclusions made
516 based on the analysis of the other biochemical blood mark-
517 ers. Indicators for assessing LPO included I_{max} (maximum
518 intensity, which reflects the potential ability of a biological
519 object to free radical oxidation) and S (the so-called "light
520 sum", reflects the content of RO_2 radicals corresponding to
521 the termination of the free radical oxidation chain). In acute
522 hepatitis caused by IZ, there was a significant increase in
523 free radical oxidation (S, $S_1 \text{max}$, α , Z, $Z_1 \text{max}$) compared
524 with intact control, while the values of tg_2 and Dec did not
525 change significantly (Table 2).

526 Changes in the values of LPO parameters were more pro-
527 nounced in relation to the action of the studied molecules
528 than the previously described biochemical parameters. In

529 particular, all the three compounds contributed to a signifi-
530 cant decrease in the values of LPO, which increased when
531 playing the IZ model (S, $S_1 \text{max}$, α , Z, $Z_1 \text{max}$), which indi-
532 cates the antioxidant activity of VB12 derivatives (Table 2).
533 At the same time, in the 5th group (ACm), there was a more
534 pronounced decrease in the values of S and $S_1 \text{max}$ com-
535 pared to group 3 (VB12). Thus, the studied compounds
536 reduced LPO caused by IZ and increased the body's anti-
537 oxidant resource.

Histological results of VB12 derivatives against isoniazid intoxication

540 When VB12 was taken by animals in which the IZ intoxi-
541 cation was reproduced, moderately pronounced plethora of
542 portal veins was noted in the samples of liver tissue with
543 anemia of the central veins and sinusoids along with moder-
544 ately pronounced lymphohistiocytic infiltration of the portal
545 tract stroma with the presence of single eosinophils in the
546 infiltrate (Fig. 6A). Damage to hepatocytes was observed in

Table 2 Intensity indicators of the lipid peroxidation process in blood plasma in the studied groups

Group No	Groups	Indicators							
		I_{max} , mV	S, mV•sec	S_1 max, mV•sec	α	Z, sec	Z_1 max, sec	tg_2 , mV/sec	Dec
1	Control intact	53.6 ± 6.3	1783 ± 197	733 ± 134	0.25 ± 0.06	14.8 ± 3.8	13.2 ± 4.1	-14.4 ± 3.1	-0.54 ± 0.47
2	IZ	72.6 ± 12.1 ^a	773 ± 136 ^a	1723 ± 436 ^a	0.41 ± 0.10 ^a	24.8 ± 6.0 ^a	23.9 ± 6.1 ^a	-13.5 ± 3.8	-0.27 ± 0.12
3	IZ + VB12	86.0 ± 15.2	1293 ± 128 ^b	1233 ± 124 ^b	0.26 ± 0.05 ^b	15.4 ± 2.7 ^b	14.7 ± 2.7 ^b	-18.9 ± 6.3	-0.36 ± 0.10
4	IZ + AQ	83.0 ± 25	1199 ± 288 ^b	1136 ± 278 ^b	0.25 ± 0.05 ^b	14.8 ± 3.0 ^b	14.1 ± 3.0 ^b	-19.5 ± 7.7	-0.39 ± 0.17
5	IZ + ACm	76.0 ± 22.4	1133 ± 134 ^{b,c}	1081 ± 126 ^{b,c}	0.26 ± 0.08 ^b	15.9 ± 4.7 ^b	15.2 ± 4.7 ^b	-17.7 ± 6.4	-0.37 ± 0.12

Data are in the format of mean ± SD (standard deviation). Significant differences were noted: ^abetween intact (group 1) and model (group 2); ^bbetween the control group (group 2, IZ) and comparison groups (3, 4, 5); ^cbetween VB12 (group 3) and groups 4, 5 (Mann–Whitney test); $P < 0.05$

ACm aquacyanocobyrinic acid heptamethyl ester

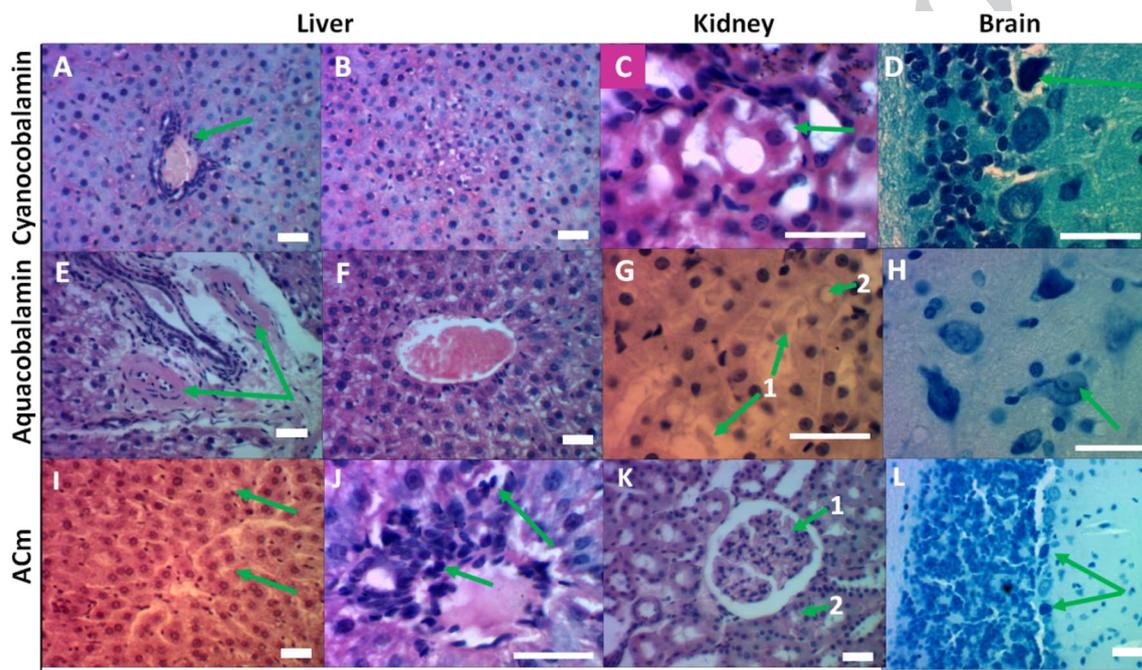


Fig. 6 Histological analysis of the studied compounds against IZ intoxication: VB12 (A–D), AQ (E–H), and ACm (I–L). Staining with hematoxylin and eosin (A, B, C, E, F, I, J, K), PAS-reaction (G), Nissl toluidine blue (D, H, L). Magnification ×480 (A, B, E, F, I, K, L) and ×1200 (C, D, G, H, J); scale bar corresponds to 100 μm. **A** Congestion of the portal vein, inflammatory infiltrate with single eosinophils. **B** Vacuolar dystrophy of hepatocytes in the center of the hepatic lobule. **C** Vacuolar degeneration of nephrocytes of the proximal convoluted tubule. **D** Hyperchromia, wrinkling of the piriform neuron of the cerebellum. **E** Spastic state of the hepatic artery

branches against the background of mild inflammatory infiltration of the portal tract stroma. **F** Plethora of the central vein, vacuolar degeneration of hepatocytes in the center of the lobule. **G** In the lumen of the distal convoluted tubules, insoluble homogeneous masses (1), focal vacuolar degeneration of nephrocytes (2). **H** Swelling of the pyramidal neuron with vacuolization of the cytoplasm. **I** Focal vacuolar degeneration of hepatocytes. **J** Single eosinophils in the composition of the inflammatory infiltrate. **K** Functional plethora of the glomerulus (1), focal degeneration of nephrocytes (2). **L** Pycnosis, hyperchromia of pear-shaped neurons of the cerebellum

547 the central zone of the hepatic lobules and was expressed as
548 vacuolar degeneration (Fig. 6B) and diffuse focal intracel-
549 lular cholestasis. In the cortical substance of the kidneys,
550 acute venous plethora was noted, vacuolar degeneration of
551 nephrocytes of the proximal convoluted tubules was focal in
552 nature (Fig. 6C). In the brain of animals of this group, the
553 arteries were in a state of moderately pronounced spasm,

venous plethora persists against the background of perivas- 554
cular edema of the nervous tissue. Single pyramidal and 555
pear-shaped neurons were noted in the state of pycnosis, 556
while the nuclei and organelles of the cytoplasm were pre- 557
served (Fig. 6D). The integral expert score of histological 558
damage was 3.2 ± 1.5 for IZ model and 2.3 ± 1.0 for VB12 559
which was borderline significant ($P = 0.056$). 560

561 AQ (group 4): in most cases, plethora of central veins
562 and sinusoids of the precentral zone of the liver was noted,
563 while in two cases, spasm of the hepatic arteries was noted
564 (Fig. 6E). Vacuole degeneration of hepatocytes in the centers
565 of the hepatic lobules was subtotal (Fig. 6F). The severity
566 of the stromal inflammatory infiltrate was relatively low. In
567 the kidneys, there was a moderately pronounced plethora
568 of the cortical and medulla. In the lumens of the convo-
569 luted tubules, PAS-negative masses were determined while
570 vacuolar degeneration of nephrocytes was focal (Fig. 6G).
571 In the brain, against the background of hemostasis in the
572 microcirculatory bed, a moderately pronounced perivascular
573 edema of the nervous tissue was observed. Neuronal inju-
574 rries differed from other groups and were characterized by
575 focal swelling with cytoplasmic vacuolization (Fig. 6H). The
576 integral expert score of histological damage for AQ was 2.4
577 ± 1.1 ($P = 0.058$).

578 In the case of ACm (group 5), in liver, the blood filling of
579 all parts of the vascular bed was in most of samples. Degen-
580 eration of a few hepatocytes was noted, and, in general, the
581 histoarchitectonics of the hepatic lobule was not disturbed
582 (Fig. 6I). Lymphohistiocytic infiltration of the stroma in the
583 portal tracts was mild, with only single eosinophils in its
584 composition (Fig. 6J). The cortical substance of the kidneys
585 had normal blood supply; vacuolar degeneration was seen
586 only in single nephrocytes (Fig. 6K). In some samples, the
587 lumens of the distal tubules were obturated with insoluble
588 homogeneous masses. In the brain, signs of moderately pro-
589 nounced plethora, edema of the nervous tissue, focal damage
590 to the neurons of the cortex and cerebellum by the type of
591 hyperchromia and pycnosis remained (Fig. 6L). The integral
592 expert score of histological damage for ACm was 2.0 ± 1.4
593 ($P < 0.05$).

594 Thus, these VB12 compounds minimized the level of
595 damage to hepatocytes and the severity of inflammatory
596 cell infiltration. The corrin substances studied prevented the
597 development of acute ischemia of the renal cortex, reduced
598 the level of nephrocyte dystrophy, although they did not
599 contribute to maintaining the normal functioning of the
600 glomerular filter (especially ACm). The level of structural
601 brain damage associated with the use of IZ remained the
602 same for all substances studied. In the AQ group, the nature
603 of neuronal damage was characterized by swelling of nerve
604 cells, while pycnotic changes predominated in other groups.
605 According to histological data, the most pronounced hepato-
606 protective effect was observed for ACm.

607 Conclusion

608 This study provides the first data on the antidote effect of a
609 semi-synthetic derivative of VB12 (ACm) compared with
610 VB12 and AQ in relation to IZ, an anti-tuberculosis drug

used in first-line therapy. In vitro experiments showed that
both protonated and deprotonated forms of IZ can react
with ACm. Deprotonation of IZ can lead to an increase in
the reduction rate of ACm, which is the oxidizing agent of
IZ in aqueous solution at various pH values. Analysis of
the kinetic data showed that oxidation proceeds through
complexation between ACm and IZ. This is followed by a
rapid electron transfer within the corrin core to produce the
reduced form of ACm and a hydrazyl radical, which further
transforms into end non-toxic products. It is assumed that
ACm reduces the formation of toxic metabolites from IZ
oxidation.

In vivo experiments revealed that acute IZ intoxication in
rats resulted in elevated levels of AST and of direct bilirubin,
with a decrease in total protein levels in the blood. Damage
to the liver, kidneys, and brain were confirmed histologi-
cally. Antidote usage of VB12 and ACm, which is currently
not included in the official list of biologically active deriva-
tives of VB12, contributed to the normalization of AST lev-
els, of AQ and ACm—to the normalization of total protein in
the blood serum. ACm, in addition to normalizing the levels
of protein and AST, also appears to normalize the level of
bilirubin in the blood. All studied compounds significantly
reduced LPO. A more pronounced decrease in the values
of S and S₁max in LPO was noted for ACm compared with
VB12. Histological analysis confirmed the protective effects
of the substances studied not only on the liver tissue, but also
on the kidneys and brain. The use of all compounds mini-
mized the level of damage to hepatocytes and the severity of
inflammatory cell infiltration, prevented the development of
acute ischemia in the renal cortex, and reduced the degen-
eration of nephrocytes and neurons. According to histology,
the most pronounced hepatoprotective effect was observed
for ACm.

The results suggest that ACm can serve as an antidote
for IZ poisoning. A direct reaction of the compound with
a toxicant is possible, leading to the formation of low-toxic
derivatives. A more pronounced effect on hepatoprotection
of ACm can also be explained by the fact that hydrophobic
substances are better accumulated in the liver than hydro-
philic ones (VB12 and AQ). In addition, in contrast to the
data on ACm obtained in the first part of this work, VB12
and AQ are not able to oxidize IZ (Petrova et al. 2014), but
AQ can bind it (Gromova et al. 2022). VB12 does not react
with IZ. The positive effect of VB12 and its aqueous form is
explained by the fact that they have a high biological activ-
ity, enhance tissue regeneration, including higher liver func-
tion. The results obtained in this work showed the feasibility
of further study of VB12 and its derivatives, with special
attention to its semi-synthetic hydrophobic derivatives. The
conducted study indicates the prospect of correcting vitamin
therapy by adding VB12 to isoniazid pharmacotherapy in
patients diagnosed with tuberculosis.

664 **Acknowledgements** This work was supported by Ministry of Science
665 and Higher Education of the Russian Federation FZZW-2023-0009 in
666 the part of spectral studies.

667 **Author contributions** All authors equally contributed to this work.
668 Unfortunately, Oscar I. Koifman passed away on 31/12/2023.

669 **Funding** Russian Science Foundation, 20-12-00175p, Olga A. Gro-
670 mova, Ministry of Science and Higher Education of the Russian Fed-
671 eration, FZZW-2020-0008, Olga A. Gromova.

672 **Data availability** There are no new datasets generated in this manu-
673 script. Data are available upon reasonable request.

674 **Code availability** Not applicable.

675 Declarations

676 **Conflict of interest** The authors declare no conflict of interest.

677 **Ethical approval** Not applicable.

678 **Consent for publication** Not applicable.

679 **Consent to participate** Not applicable.

680 References

- 681 Ariga K, Mori T, Nakanishi W, Hill JP (2017) Solid surface vs liquid
682 surface: nanoarchitectonics, molecular machines, and DNA ori-
683 gami. *Phys Chem Chem Phys* 19:23658–23676. [https://doi.org/](https://doi.org/10.1039/C7CP02280H)
684 [10.1039/C7CP02280H](https://doi.org/10.1039/C7CP02280H)
- 685 Ariga K, Nishikawa M, Mori T, Takeya J, Shrestha LK, Hill JP (2019)
686 Self-assembly as a key player for materials nanoarchitectonics.
687 *Sci Technol Adv Mater* 20:51–95. [https://doi.org/10.1080/14686](https://doi.org/10.1080/14686996.2018.1553108)
688 [996.2018.1553108](https://doi.org/10.1080/14686996.2018.1553108)
- 689 Ariga K, Tsai KC, Shrestha LK, Hsu SH (2021) Life science nanoarchi-
690 tectonics at interfaces. *Mater Chem Front* 5:1018–1032. [https://](https://doi.org/10.1039/D0QM00615G)
691 doi.org/10.1039/D0QM00615G
- 692 ASPCA Animal Poison Control Center Phone Number: (888) 426–
693 4435, [https://www.aspcapro.org/topics-animal-health/toxicology-](https://www.aspcapro.org/topics-animal-health/toxicology-poison-control)
694 [poison-control](https://www.aspcapro.org/topics-animal-health/toxicology-poison-control), Accessed 26 May 2022.
- 695 Badrinath M, John S. Isoniazid Toxicity. 2022 Jun 27. In: StatPearls
696 [Internet]. Treasure Island (FL): StatPearls Publishing; 2022 Jan–.
697 PMID: 30285383, Bookshelf ID: [NBK531488](https://www.ncbi.nlm.nih.gov/books/NBK531488). [https://www.ncbi.](https://www.ncbi.nlm.nih.gov/books/NBK531488)
698 [nlm.nih.gov/books/NBK531488](https://www.ncbi.nlm.nih.gov/books/NBK531488)
- 699 Brenner W, Ronson TK, Nitschke JR (2017) Separation and selective
700 formation of fullerene adducts within an MII 8L6 cage. *J Am*
701 *Chem Soc* 139:75–78. <https://doi.org/10.1021/jacs.6b11523>
- 702 Brown MJ, Symonowicz C, Medina LV, Bratcher NA, Buckmaster
703 CA, Klein H, Anderson LC (2017) Culture of care: organizational
704 responsibilities. Management of animal care and use programs in
705 research, education, and testing, 11–26.
- 706 Couto M, Cates C (2019) Laboratory guidelines for animal care. Ver-
707 tebrate embryogenesis. Humana, New York, NY, pp 407–430.
708 https://doi.org/10.1007/978-1-4939-9009-2_25
- 709 Dale WE, Haluska GJ, Horne D (2018) Program documentation and
710 monitoring. In: Weichbrod RH, Thompson GA, Norton JN (eds)
711 Management of animal care and use programs in research, edu-
712 cation, and testing, 2nd edn. CRC Press/Taylor & Francis, Boca
713 Raton (FL). <https://doi.org/10.1201/9781315152189> (Chapter 15.
714 PMID: 29787191)

- de Moraes Profirio D, Pessine FBT (2018) Formulation of func-
tionalized PLGA nanoparticles with folic acid-conjugated chi-
tosan for carboplatin encapsulation. *Eur Polym J* 108:311–321.
<https://doi.org/10.1016/j.eurpolymj.2018.09.011>
- Edgar A, Koh M, Patel R (2022) A case of accidental isoniazid over-
dose presenting with nonspecific symptoms. *Cureus*. <https://doi.org/10.7759/cureus.23218>
- Erwin ER, Addison AP, John SF, Olaleye OA, Rosell RC (2019)
Pharmacokinetics of isoniazid: the good, the bad, and the alter-
natives. *Tuberculosis* 116:S66–S70. <https://doi.org/10.1016/j.tube.2019.04.012>
- Gebremicael G, Alemayehu M, Sileshi M, Geto Z, Gebreegziabxier
A, Tefera H, Desta K (2019) The serum concentration of vita-
min B12 as a biomarker of therapeutic response in tuberculosis
patients with and without human immunodeficiency virus (HIV)
infection. *Int J Gen Med* 12:353–361. <https://doi.org/10.2147/IJGM.S218799>
- Gomes J, Durães D, Sousa A, Afonso H (2019) Isoniazid-induced
acute psychosis in a patient with pleural tuberculosis. *Case Rep*
Psychiatry. <https://doi.org/10.1155/2019/4272941>
- Gromova OA, Torshin IY, Maiorova LA, Koifman OI, Salsnikov DS
(2021) Bioinformatic and chemoneurocytological analysis of
the pharmacological properties of vitamin B12 and some of
its derivatives. *J Porphyrins Phthalocyanines* 25(09):835–842.
<https://doi.org/10.1142/S1088424621500644>
- Gromova OA, Maiorova LA, Salsnikov DS, Demidov VI, Kalacheva
AG, Torshin IY, Koifman OI (2022) Vitamin B₁₂ hydrophobic
derivative exhibits bioactivity: biomedical and photophysical
study. *BioNanoScience* 12(1):74–82. <https://doi.org/10.1007/s12668-021-00916-4>
- Guéant JL, Guéant-Rodriguez RM, Oussalah A, Zuily S, Rosenberg
I (2023) Hyperhomocysteinemia in cardiovascular diseases:
revisiting observational studies and clinical trials. *Thromb*
Haemost 123:270–282. <https://doi.org/10.1055/a-1952-1946>
- Haloul M, Vinjamuri SJ, Naquiallah D, Mirza MI, Maryam Q, Chan-
dra H, Abeer MM (2020) Hyperhomocysteinemia and low folate
and vitamin B12 are associated with vascular dysfunction and
impaired nitric oxide sensitivity in morbidly obese patients.
Nutrients 12:2014. <https://doi.org/10.3390/nu12072014>
- Hendry-Hofer TB, Ng PC, McGrath AM, Soules K, Mukai DS, Chan
A, Bebartha VS (2021) Intramuscular cobinamide as an antidote
to methyl mercaptan poisoning. *Inhal Toxicol* 33(1):25–32.
<https://doi.org/10.1080/08958378.2020.1866123>
- Jiang J, Chan A, Ali S, Saha A, Haushalter KJ, Lam WLM, Boss GR
(2016) Hydrogen sulfide—mechanisms of toxicity and devel-
opment of an antidote. *Sci Rep* 6(1):1–10. <https://doi.org/10.1038/srep20831>
- Kharitonova NV, Maiorova LA, Koifman OI (2018) Aggregation
behavior of unsubstituted magnesium porphyrine in mon-
olayers at air–water interface and in Langmuir–Schaefer films.
J Porphyr Phthalocyanines 22:509–520. <https://doi.org/10.1142/S1088424618500505>
- Lee J, Mahon SB, Mukai D, Burney T, Katebian BS, Chan A, Bren-
ner M (2016) The vitamin B12 analog cobinamide is an effective
antidote for oral cyanide poisoning. *J Med Toxicol* 12(4):370–
379. <https://doi.org/10.1007/s13181-016-0566-4>
- Levy J, Rodriguez-Guéant RM, Oussalah A, Jeannesson E, Wahl
D, Ziuly S, Guéant JL (2021) Cardiovascular manifestations of
intermediate and major hyperhomocysteinemia due to vitamin
B12 and folate deficiency and/or inherited disorders of one-
carbon metabolism: a 3.5-year retrospective cross-sectional
study of consecutive patients. *Am J Clin Nutr*. 113:1157–1167.
<https://doi.org/10.1093/ajcn/nqaa432>
- Maiorova LA, Koifman OI, Burmistrov VA, Kuvshinova SA,
Mamontov AO (2015) 2D M-nanoaggregates in Langmuir

715
716
717
718
719
720
721
722
723
724
725
726
727
728
729
730
731
732
733
734
735
736
737
738
739
740
741
742
743
744
745
746
747
748
749
750
751
752
753
754
755
756
757
758
759
760
761
762
763
764
765
766
767
768
769
770
771
772
773
774
775
776
777
778
779

- 780 layers of calamite mesogen. *Prot Metals and Phys Chem Surf*
781 51(1):85–92. <https://doi.org/10.1134/S2070205115010074>
- 782 Maiorova LA, Kobayashi N, Zyablov SV, Bykov VA, Nesterov SI,
783 Kozlov AV, Koifman OI (2018) Magnesium porphine supermol-
784 ecules and two-dimensional nanoaggregates formed using the
785 Langmuir-Schaefer technique. *Langmuir* 34:9322–9329. <https://doi.org/10.1021/acs.langmuir.8b00905>
- 786
787 Maiorova LA, Kobayashi N, Salnikov DS, Kuzmin SM, Basova TV,
788 Koifman OI, Yang P (2023) Supermolecular nanoentities of
789 vitamin B 12 derivative as a link in the evolution of the par-
790 ent molecules during self-assembly at the air–water interface.
791 *Langmuir* 39:3246–3254. <https://doi.org/10.1021/acs.langmuir.2c02964>
- 792
793 Maiorova LA, Gromova OA, Torshin IYu, Bukreeva TV, Pallaeva TN,
794 Nabatov BV, Yabbarov NG (2024) Nanoparticles of nucleotide-
795 free analogue of vitamin B12 formed in protein nanocarriers and
796 their neuroprotective activity in vivo. *Colloids Surf B Biointer-*
797 *faces* 244:114165. <https://doi.org/10.1016/j.colsurfb.2024.114165>
- 798
799 Martin SJ, Prince SE (2017) Comparative modulation of levels of oxi-
800 dative stress in the liver of anti-tuberculosis drug treated wistar
801 rats by vitamin B12, beta-carotene, and spirulinafusiformis: role
802 of NF- κ B, iNOS, IL-6, and IL-10. *J Cell Biochem* 118:3825–
803 3833. <https://doi.org/10.1002/jcb.26032>
- 804
805 Morris AL, Mohiuddin SS. *Biochemistry, nutrients*. 2022 May 8. In:
806 *StatPearls* [Internet]. Treasure Island (FL): StatPearls Publish-
807 ing; 2022 Jan–. PMID: 32119432, <https://www.ncbi.nlm.nih.gov/books/NBK554545>
- 808
809 Nowak M, Brown TD, Graham A, Helgeson ME, Mitragotri S (2020)
810 Size, shape, and flexibility influence nanoparticle transport across
811 brain endothelium under flow. *Bioeng Transl Med* 5:e10153.
812 <https://doi.org/10.1002/btm2.10153>
- 813
814 O'Connor C, Brady MF. *Isoniazid* 2022 Apr 8. In: *StatPearls* [Internet].
815 Treasure Island (FL): StatPearls Publishing; 2022 Jan. PMID:
816 32491549. <https://www.ncbi.nlm.nih.gov/books/NBK557617>.
- 817
818 Oh C, Keats EC, Bhutta ZA (2020) Vitamin and mineral supplementa-
819 tion during pregnancy on maternal, birth, child health and devel-
820 opment outcomes in low- and middle-income countries: a system-
821 atic review and meta-analysis. *Nutrients* 12(2):491. <https://doi.org/10.3390/nu12020491>. (PMID:32075071;PMCID:PMC7071347)
- 822
823 Oldacre AN, Friedman AE, Cook TR (2017) A self-assembled cofacial
824 cobalt porphyrin prism for oxygen reduction catalysis. *J Am Chem*
825 *Soc* 139:1424–1427. <https://doi.org/10.1021/jacs.6b12404>
- 826
827 Orssaud C, Nguyen DT, Rouzard C, Pavie J, Pinot J, Lortholary O,
828 Robert MP (2022) Dépistage et prévention des neuropathies opti-
829 questoxiques aux anti-mycobactériens: proposition de recomman-
830 dations. *J Français d'ophtalmologie* 45(5):495–503. <https://doi.org/10.1016/j.jfo.2021.08.016>
- 831
832 Ouyang M, Du Y, Meng F, Zhang X, Zhuang Q, Ma Y, Liu H, Pang
833 M, Cai T, Cai Y (2019) Polymer-lipid hybrid nanoparticles: a
834 novel drug delivery system for enhancing the activity of Psoralen
835 against breast cancer. *Int J Pharm* 561:274–282. <https://doi.org/10.1016/j.ijpharm.2019.03.006>
- 836
837 Petrova MV, Maiorova LA, Bulkina TA, Ageeva TA, Koifman OI,
838 Gromova OA (2014) Nanostructure of zinc(II) tetraphenylpor-
839 phyrinate Langmuir M-monolayers formed with diluted solu-
840 tion. *Macromolecules* 47(3):267–271. <https://doi.org/10.1021/acs.macromol.4b00606>
- 841
842 Philipopoulos GP, Tat J, Chan A, Jiang J, Mukai D, Burney T, Boss
843 GR (2022) Methyl mercaptan gas: mechanisms of toxicity and
844 demonstration of the effectiveness of cobinamide as an antidote
845 in mice and rabbits. *Clin Toxicol* 60(5):615–622. <https://doi.org/10.1080/15563650.2021.2017949>
- 846
847 Soares S, Sousa J, Pais A, Vitorino C (2018) Nanomedicine: principles,
848 properties, and regulatory issues. *Front Chem* 6:360. <https://doi.org/10.3389/fchem.2018.00360>
- 849
850 Sridhar A, Sandeep Y, Krishnakishore C, Sriramnaveen P, Manjusha Y,
851 Sivakumar V (2012) Fatal poisoning by isoniazid and rifampicin.
852 *Indian Nephrol* 22(5):385. <https://doi.org/10.4103/0971-4065.103930>
- 853
854 Stulz E (2017) Nanoarchitectonics with porphyrin functionalized DNA.
855 *Acc Chem Res* 50:823–831. <https://doi.org/10.1021/acs.accounts.6b00583>
- 856
857 Tumakov SO, Dereven'kov IA, Sal'nikov DS, Makarov SV (2019)
858 Kinetics of the reaction between cobinamide and isoniazid in
859 aqueous solutions. *Russ J Phys Chem A* 93(2):265–270. <https://doi.org/10.1134/S0036024419020274>
- 860
861 Valkova LA, Shabyshev LS, Feigin LA, Akopova OB (1996a) For-
862 mation and X-ray diffraction investigation of Langmuir-Blodgett
863 films of liquid crystalline substituted crown esters. *Mol Cryst Liq*
864 *Cryst Sci Technol* 6:291–298
- 865
866 Valkova L, Betrencourt C, Hochapfel A, Myagkov IV, Feigin LA
867 (1996b) Monolayer study of monensin and lasalocid in the gas
868 state. *Mol Cryst Liq Cryst Sci Technol A Mol Cryst Liq Cryst*
869 287:269–273. <https://doi.org/10.1080/10587259608038763>
- 870
871 Valkova LA, Shabyshev LS, Feigin LA, Akopova OB (1997) Prepara-
872 tion and X-ray study of Langmuir-Blodgett films of liquid crys-
873 tal 4,5'-bis(4-decyloxybenzoyloxybenzylideneamino) dibenzo-
874 18-crown-6. *Izv Akad Nauk, Ser Fiz* 61:631–636 (WOS:
875 A1997WZ50000048)
- 876
877 Valkova LA, Shabyshev LS, Borovkov NYu, Feigin LA, Rustichelli F
878 (1999) Supramolecular assembly formation in monolayers of tert-
879 butyl substituted copper phthalocyanine and tetrabenzotriazaporph-
880 in. *J Incl Phenom Macrocycl Chem* 35:243–249. <https://doi.org/10.1023/A:1008147031935>
- 881
882 Valkova L, Borovkov N, Maccioni E, Pisani M, Rustichelli F, Erokhin
883 V, Nicolini C (2002) Influence of molecular and supramolecular
884 factors on sensor properties of Langmuir-Blodgett films of tert-
885 butyl-substituted copper azaporphyrines towards hydrocarbons.
886 *Colloids Surf A Physicochem Eng Asp* 198–200:891–896. [https://doi.org/10.1016/S0927-7757\(01\)01016-0](https://doi.org/10.1016/S0927-7757(01)01016-0)
- 887
888 Vilchêze C, Jacobs WR Jr (2019) The isoniazid paradigm of killing,
889 resistance, and persistence in mycobacterium tuberculosis. *J Mol*
890 *Biol* 431(18):3450–3461. <https://doi.org/10.1016/j.jmb.2019.02.016>
- 891
892 Villar D, Knight MK, Holding J, Barret GH, Buck WB (1995) Treat-
893 ment of acute isoniazid overdose in dogs. *Vet Hum Toxicol*
894 37(5):473–477
- 895
896 Vu TT, Maiorova LA, Berezin DB, Koifman OI (2016) Formation
897 and study of nanostructured M-monolayers and LS-films of triphen-
898 ylcorrole. *Macromolecules* 49:73–79. <https://doi.org/10.1021/acs.macromol.5b01205>
- 899
900 Wang J, Zhou Y, Zhao C, Xiong K, Liu Y, Zhao S, Ma A (2024)
901 Dietary patterns and the risk of tuberculosis-drug-induced liver
902 injury: a cohort study. *Front Nutr* 11:1393523. <https://doi.org/10.3389/fnut.2024.1393523>
- 903
904 Webre WA, Gobeze HB, Shao S, Karr PA, Ariga K, Hill JP, D'Souza F
905 (2018) Fluoride-ion binding promoted photoinduced charge separa-
906 tion in a self-assembled C60 alkyl cation bound bis-crown ether-
907 oxoporphyrinogen supramolecule. *Chem Commun* 54:1351–1354.
908 <https://doi.org/10.1039/C7CC09524D>
- 909
910 Xia XS, Li X, Wang L, Wang J-Z, Ma J-P, Wu C-J (2014) Supple-
911 mentation of folic acid and vitamin B12 reduces plasma levels
912 of asymmetric dimethylarginine in patients with acute ischemic
913 stroke. *J Clin Neurosci* 21:1586–1590. <https://doi.org/10.1016/j.jocn.2013.11.043>
- 914
915 Zeytunluoglu A, Arslan I (2022) Current perspectives on nanoemul-
916 sions in targeted drug delivery, In: *Handbook of research on nanoemul-*
917 *sion applications in agriculture, food, health, and*

912 biomedical sciences 118–140, [https://doi.org/10.4018/978-1-](https://doi.org/10.4018/978-1-7998-8378-4.ch006)
 913 [7998-8378-4.ch006](https://doi.org/10.4018/978-1-7998-8378-4.ch006).
 914 Zhang K, Zhu S, Li J, Jiang T, Feng L, Pei J, Liu B (2021) Targeting
 915 autophagy using small-molecule compounds to improve potential
 916 therapy of Parkinson's disease. *Acta Pharm Sin B* 11(10):3015–
 917 3034. <https://doi.org/10.1016/j.apsb.2021.02.016>
 918 Zhang T-P, Li R, Wang L-J, Tang F, Li H-M (2022) Clinical relevance
 919 of vitamin B12 level and vitamin B12 metabolic gene variation
 920 in pulmonary tuberculosis. *Fron Immunol* 13:947897. [https://doi-](https://doi.org/10.3389/fimmu.2022.947897)
 921 [org/10.3389/fimmu.2022.947897](https://doi.org/10.3389/fimmu.2022.947897)

Publisher's Note Springer Nature remains neutral with regard to
 jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds
 exclusive rights to this article under a publishing agreement with the
 author(s) or other rightsholder(s); author self-archiving of the accepted
 manuscript version of this article is solely governed by the terms of
 such publishing agreement and applicable law.

Authors and Affiliations

Olga A. Gromova¹  · Larissa A. Maiorova^{1,2}  · Denis S. Salnikov²  · Ivan Yu Torshin¹  · Vadim I. Demidov³ ·
 Irina K. Tomilova³ · O. I. Koifman² · Alla G. Kalacheva³ · Tatiana E. Bogacheva³ · Elena L. Alexakhina³ ·
 Tatiana R. Grishina³ · Andrei N. Gromov¹ · Elham Assadpour^{4,5}  · Tolulope J. Ashaolu^{6,7}  · Seid Mahdi Jafari^{8,9} 

✉ Olga A. Gromova
 unesco.gromova@gmail.com

✉ Larissa A. Maiorova
 maiorova.larissa@gmail.com

✉ Seid Mahdi Jafari
 smjafari@gau.ac.ir

Denis S. Salnikov
 densal82@mail.ru

Ivan Yu Torshin
 tiy135@yahoo.com

Vadim I. Demidov
 13vid@mail.ru

Irina K. Tomilova
 tomilovaivanovo@mail.ru

Alla G. Kalacheva
 alla_kalacheva@mail.ru

Tatiana E. Bogacheva
 tatiana.boga4iova@yandex.ru

Elena L. Alexakhina
 alexakhina2013@yandex.ru

Tatiana R. Grishina
 farma37@bk.ru

Andrei N. Gromov
 gromlogin@gmail.com

Elham Assadpour
 assadpour1170@gmail.com

Tolulope J. Ashaolu
 tolulopejoshuaashaolu@duytan.edu.vn

¹ Federal Research Center “Computer Science and Control”
 of Russian Academy of Sciences, Moscow, Russia

² Institute of Macrocyclic Compounds, Ivanovo State
 University of Chemistry and Technology, Ivanovo, Russia

³ Ivanovo State Medical University, Ministry of Health
 of Russia, Ivanovo, Russia

⁴ Food Industry Research Co., Gorgan, Iran

⁵ Food and Bio-Nanotech International Research Center
 (Fabiano), Gorgan University of Agricultural Sciences
 and Natural Resources, Gorgan, Iran

⁶ Institute for Global Health Innovations, Duy Tan University,
 Da Nang 55000, Viet Nam

⁷ Faculty of Medicine, Duy Tan University, Da Nang 55000,
 Viet Nam

⁸ Department of Food Materials and Process Design
 Engineering, Gorgan University of Agricultural Sciences
 and Natural Resources, Gorgan, Iran

⁹ Iran Food and Drug Administration, Halal Research Center
 of IRI, Ministry of Health and Medical Education, Tehran,
 Iran

Journal:	13197
Article:	6296

Author Query Form

Please ensure you fill out your response to the queries raised below and return this form along with your corrections

Dear Author

During the process of typesetting your article, the following queries have arisen. Please check your typeset proof carefully against the queries listed below and mark the necessary changes either directly on the proof/online grid or in the 'Author's response' area provided below

Query	Details Required	Author's Response
AQ1	Please check and confirm that the authors and their respective affiliations have been correctly identified and amend if necessary.	
AQ2	Author names: Please confirm if the author names are presented accurately and in the correct sequence (given name, middle name/initial, family name). Also, kindly confirm the details in the metadata are correct.	
AQ3	Please check and confirm that the data availability section is processed correctly and amend if necessary.	
AQ4	Please check and confirm that the processed Article note is correct and amend if necessary.	